

**ESTUDIO DE GMI.
ESTUDIO PROTEÓMICO COMPARATIVO ENTRE LOS IMPLANTES
DENTALES DE GMI Y DE STRAUMANN**

Hipótesis. Estudio proteómico de la primera capa de proteínas depositada

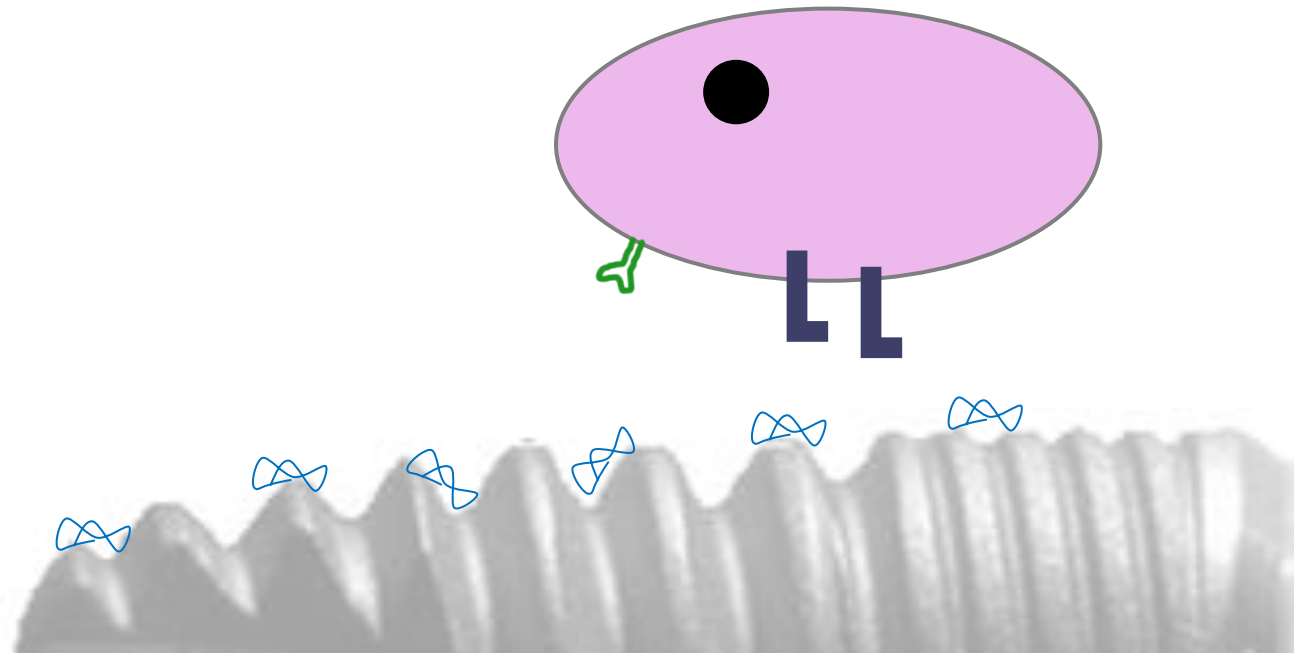
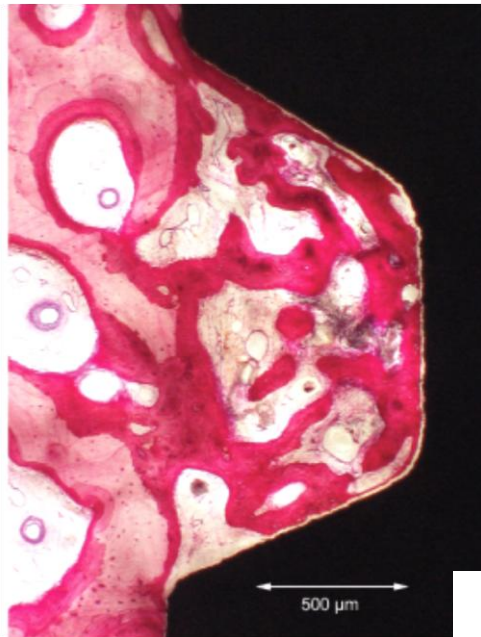
Después de la implantación: el biomaterial entra en contacto con la sangre



(en segundos)

Adsorción de proteínas de la sangre en la superficie del implante

(Andersson, 2005).



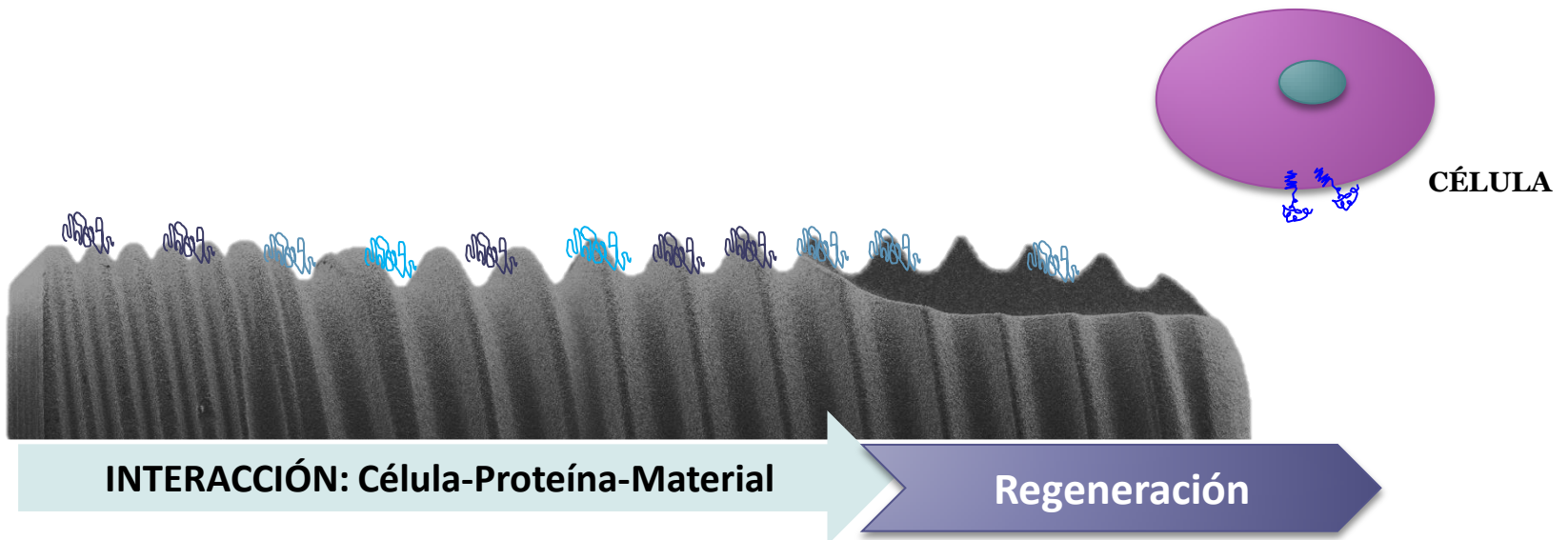
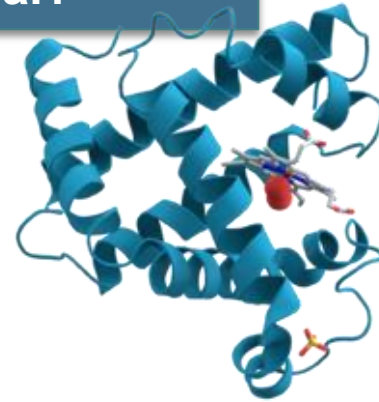
¿Por qué son importantes las proteínas adsorbidas en el biomaterial?

La primera capa de proteínas adsorbidas

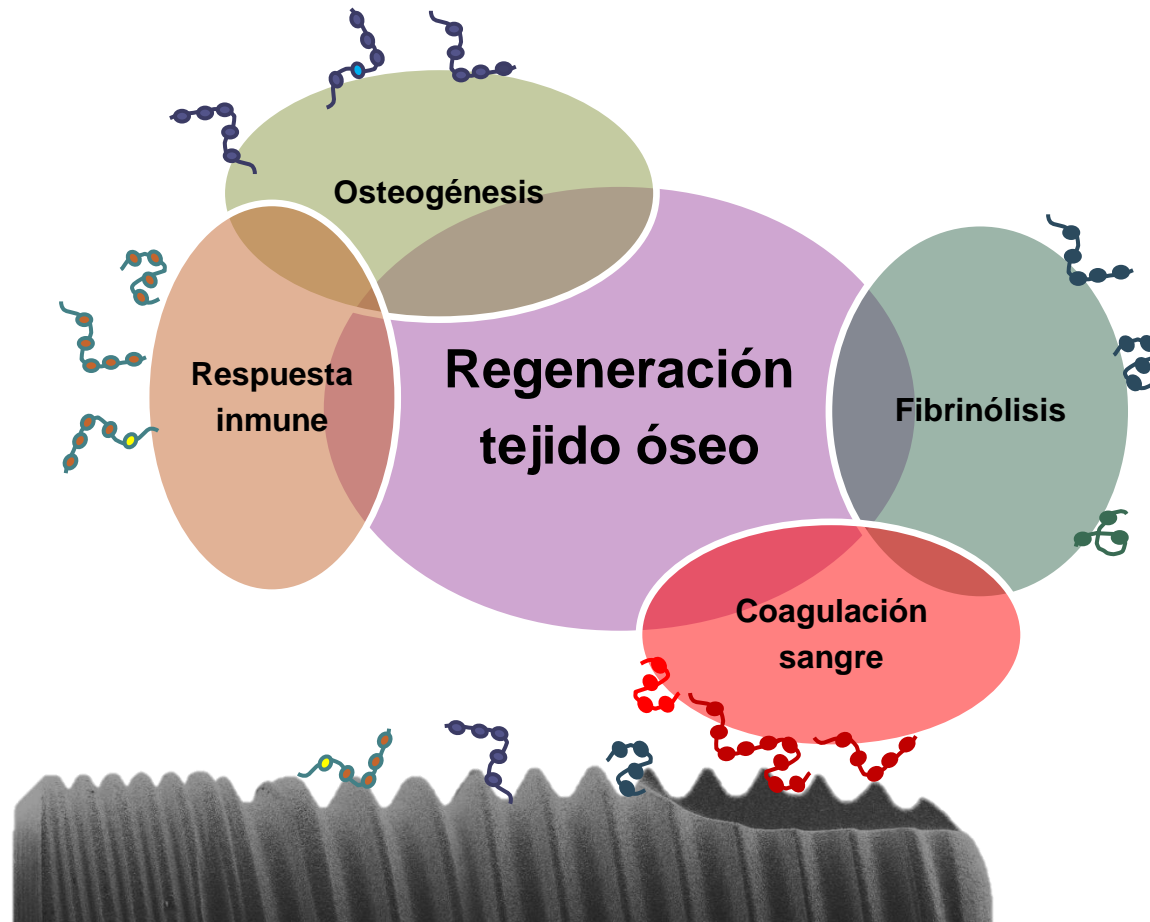


determinará la respuesta biológica al biomaterial

(Engberg, 2015)



Estudio proteómico comparativo de GMI



Proteómica: metodología

1



Incubación con suero humano (3h 37°C)

Lavados consecutivos para eliminar las proteínas no adsorbidas

H₂O Mili-Q
100mM NaCl Tris-HCl pH 7.1
N=4 (4 implantes)

Elución de las proteínas adsorbidas en el biomaterial

4% SDS, 100 mM DTT en 0.5M TEAB

2



ELUCIÓN FINAL

3

Cuantificación e identificación: Espectroscopía de masas (MS/MS)



Análisis de datos: PROGENESIS

DAVID (Database for Annotation, Visualization and Integrated Discovery)

Base de datos de familias de proteínas, útiles para la identificación y clasificación de funciones de proteínas

4



DAVID Bioinformatics Resources 6.7
National Institute of Allergy and Infectious Diseases (NIAID), NIH

ESTUDIO PROTEÓMICO COMPARATIVO :




GMI



ST





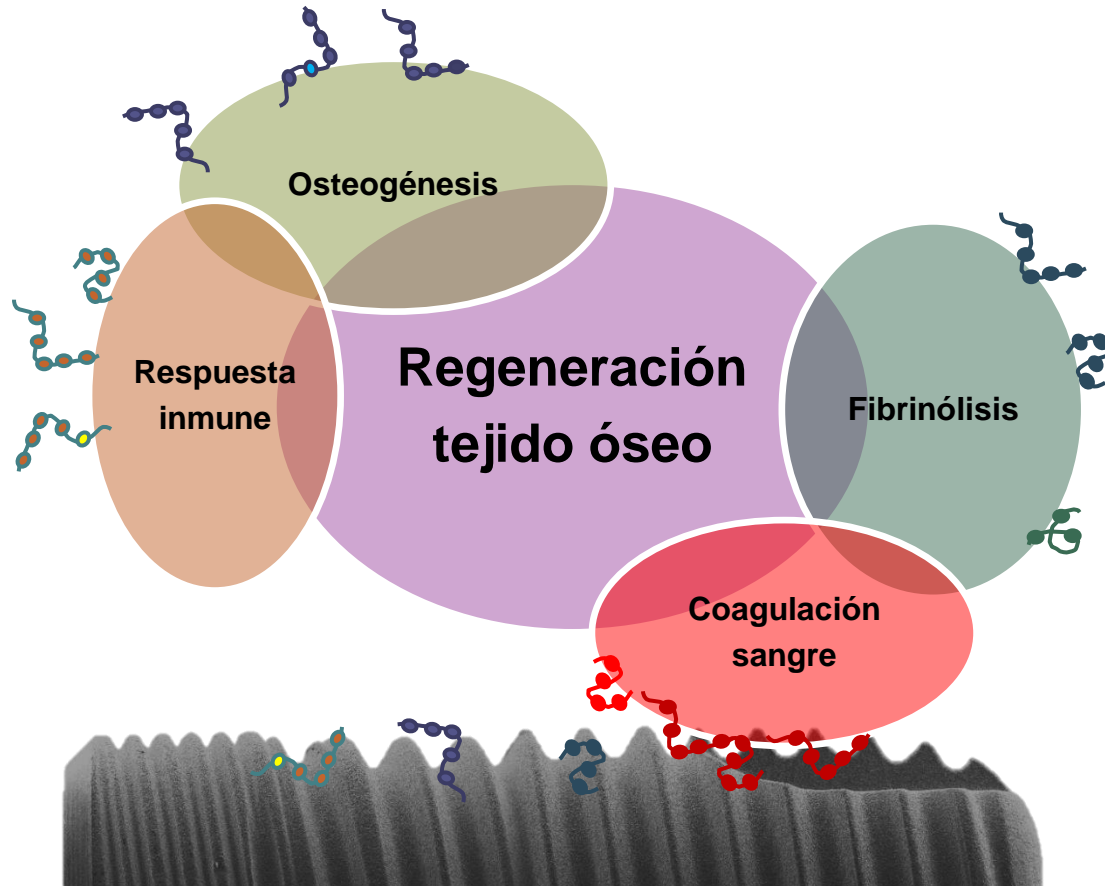
Accesión	Péptidos contados	Descripción	GMI/ST	
			Valor p	Ratio
PROC_HUMAN	8	Vitamin K-dependent protein C	1,17E-06	27,21
FA10_HUMAN	8	Coagulation factor X	1,86E-04	17,21
FA7_HUMAN	3	Coagulation factor VII	1,04E-04	16,27
TAGL2_HUMAN	2	Transgelin-2	2,40E-05	11,54
SAA1_HUMAN	2	Serum amyloid A-1 protein	1,46E-02	5,48
FA9_HUMAN	7	Coagulation factor IX	2,44E-04	5,47
APOC1_HUMAN	3	Apolipoprotein C-I	3,58E-04	5,05
FCN3_HUMAN	2	Ficolin-3	5,42E-03	4,65
CAMP_HUMAN	2	Cathelicidin antimicrobial peptide	3,93E-02	4,37
APOE_HUMAN	26	Apolipoprotein E	1,17E-03	3,61
APOC4_HUMAN	5	Apolipoprotein C-IV	2,27E-05	3,60
APOA4_HUMAN	28	Apolipoprotein A-IV	1,83E-05	3,54
CXCL7_HUMAN	6	Platelet basic protein	5,79E-03	3,05
PROS_HUMAN	13	Vitamin K-dependent protein S	2,07E-03	2,87
PLEK_HUMAN	4	Pleckstrin	1,34E-02	2,69
SEPP1_HUMAN	5	Selenoprotein P	3,31E-03	2,65
C4BPB_HUMAN	2	C4b-binding protein beta chain	4,79E-02	2,54
APOA1_HUMAN	22	Apolipoprotein A-I	2,38E-04	2,49
LBP_HUMAN	4	Lipopolysaccharide-binding protein	3,72E-02	2,38
ANT3_HUMAN	26	Antithrombin-III	6,32E-04	2,33
THRB_HUMAN	25	Prothrombin	9,76E-04	2,31
DESP_HUMAN	4	Desmoplakin	1,97E-02	2,29
FA12_HUMAN	12	Coagulation factor XII	1,15E-03	2,29
C4BPA_HUMAN	19	C4b-binding protein alpha chain	9,04E-03	2,25
FHR5_HUMAN	6	Complement factor H-related protein 5	7,01E-03	2,16
FA5_HUMAN	7	Coagulation factor V	2,80E-02	2,12
RAP1B_HUMAN	2	Ras-related protein Rap-1b	2,17E-02	2,00
CO4A_HUMAN	60	Complement C4-A	2,94E-02	1,95
CO4B_HUMAN	60	Complement C4-B	2,94E-02	1,95
GELS_HUMAN	19	Gelsolin	4,75E-04	1,94
PROP_HUMAN	3	Properdin	4,48E-02	1,87
IBP4_HUMAN	2	Insulin-like growth factor-binding protein 4	1,47E-02	1,84
APOC2_HUMAN	5	Apolipoprotein C-II	1,99E-02	1,83
MOES_HUMAN	2	Moesin	3,81E-02	1,80
PON1_HUMAN	10	Serum paraoxonase/arylesterase 1	3,35E-03	1,75
PPIB_HUMAN	2	Peptidyl-prolyl cis-trans isomerase B	2,51E-02	1,73
LIMS1_HUMAN	2	LIM and senescent cell antigen-like-containing domain protein 1	4,01E-02	1,73
CETP_HUMAN	2	Cholesteryl ester transfer protein	3,03E-02	1,53
QSOX1_HUMAN	4	Sulfhydryl oxidase 1	1,92E-02	1,50

Proteínas con menor afinidad con implantes GMI

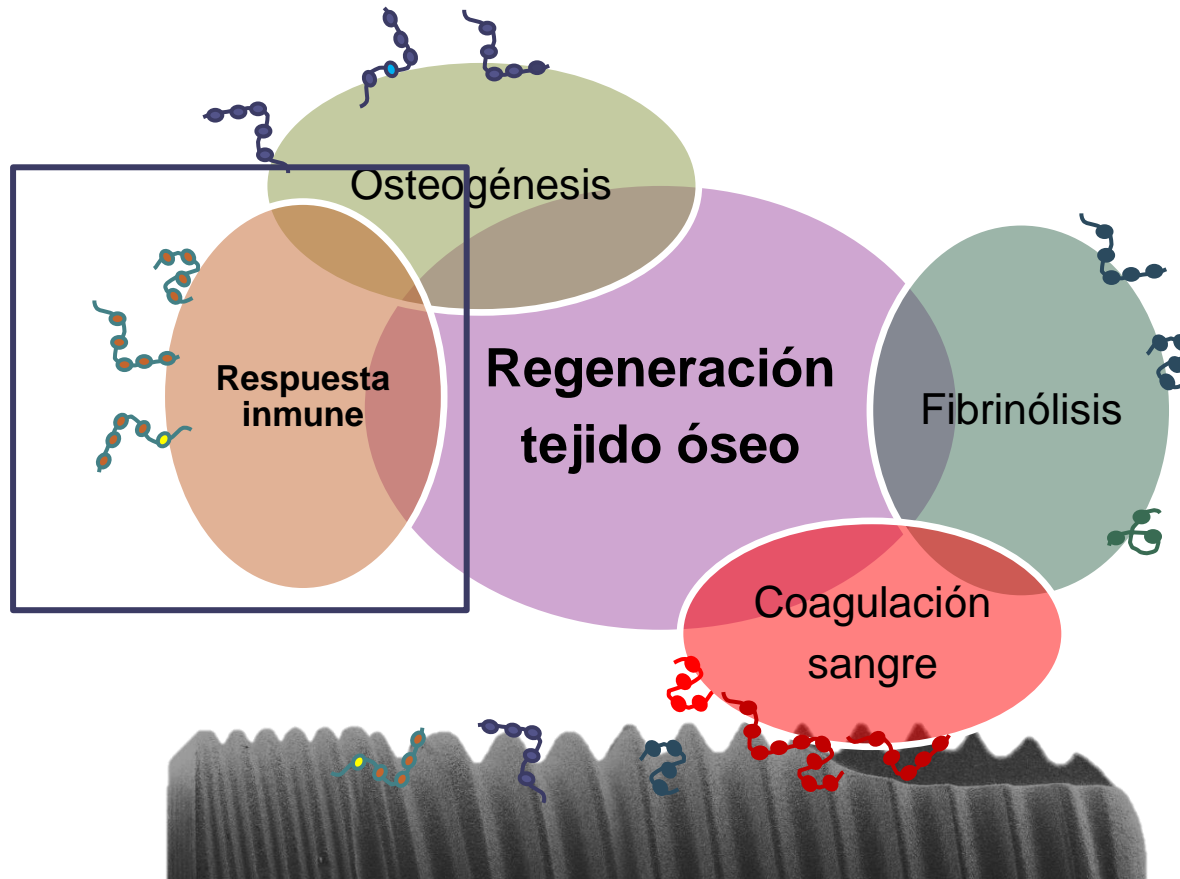


			GMI/ST	
			p value	Ratio
CO8B_HUMAN	10	Complement component C8 beta chain	3,64E-02	0,64
IGJ_HUMAN	3	Immunoglobulin J chain	4,59E-02	0,62
HV320_HUMAN	4	Ig heavy chain V-III region GAL	2,57E-02	0,62
TTHY_HUMAN	5	Transthyretin	1,70E-02	0,61
HBA_HUMAN	2	Hemoglobin subunit alpha	8,32E-03	0,53
ITIH4_HUMAN	18	Inter-alpha-trypsin inhibitor heavy chain H4	6,49E-03	0,51
CO8A_HUMAN	6	Complement component C8 alpha chain	2,48E-02	0,51
CO5_HUMAN	22	Complement C5	4,78E-02	0,49
IGHG2_HUMAN	14	Ig gamma-2 chain C region	1,38E-02	0,47
IGHG4_HUMAN	8	Ig gamma-4 chain C region	1,42E-02	0,44
FETUA_HUMAN	7	Alpha-2-HS-glycoprotein	1,08E-03	0,40
PGRP2_HUMAN	8	N-acetylmuramoyl-L-alanine amidase	1,81E-02	0,37
KNG1_HUMAN	17	Kininogen-1	6,98E-05	0,33
CFAB_HUMAN	19	Complement factor B	2,06E-02	0,33
FA11_HUMAN	20	Coagulation factor XI	6,27E-03	0,33
AMBP_HUMAN	8	Protein AMBP	5,48E-03	0,32
RET4_HUMAN	5	Retinol-binding protein	1,62E-03	0,31
VTDB_HUMAN	16	Vitamin D-binding protein	1,34E-02	0,30
IGHA1_HUMAN	11	Ig alpha-1 chain C region	1,34E-03	0,28
FCN1_HUMAN	2	Ficolin-1	7,85E-03	0,26
IGHA2_HUMAN	8	Ig alpha-2 chain C region	5,53E-03	0,24
HPT_HUMAN	21	Haptoglobin	4,07E-03	0,22
CERU_HUMAN	24	Ceruloplasmin	3,71E-04	0,22
CBG_HUMAN	2	Corticosteroid-binding globulin	5,97E-03	0,22
ANGT_HUMAN	6	Angiotensinogen	2,07E-03	0,21
AACT_HUMAN	11	Alpha-1-antichymotrypsin	4,84E-04	0,20
ITIH1_HUMAN	10	Inter-alpha-trypsin inhibitor heavy chain H1	2,50E-03	0,19
ALBU_HUMAN	57	Serum albumin	1,13E-02	0,18
A1AT_HUMAN	20	Alpha-1-antitrypsin	1,05E-03	0,17
TRFE_HUMAN	42	Serotransferrin	5,42E-03	0,15
A1AG2_HUMAN	7	Alpha-1-acid glycoprotein 2	2,30E-02	0,12
A1AG1_HUMAN	8	Alpha-1-acid glycoprotein 1	3,51E-03	0,12
ZA2G_HUMAN	11	Zinc-alpha-2-glycoprotein	7,13E-04	0,12
AFAM_HUMAN	10	Afamin	1,22E-03	0,12
A2MG_HUMAN	50	Alpha-2-macroglobulin	3,84E-03	0,11
HEMO_HUMAN	16	Hemopexin	4,91E-03	0,10
ITIH3_HUMAN	2	Inter-alpha-trypsin inhibitor heavy chain H3	1,47E-03	0,10
A1BG_HUMAN	9	Alpha-1B-glycoprotein	2,28E-03	0,09
A2GL_HUMAN	4	Leucine-rich alpha-2-glycoprotein	6,02E-03	0,08

Proceso complejo



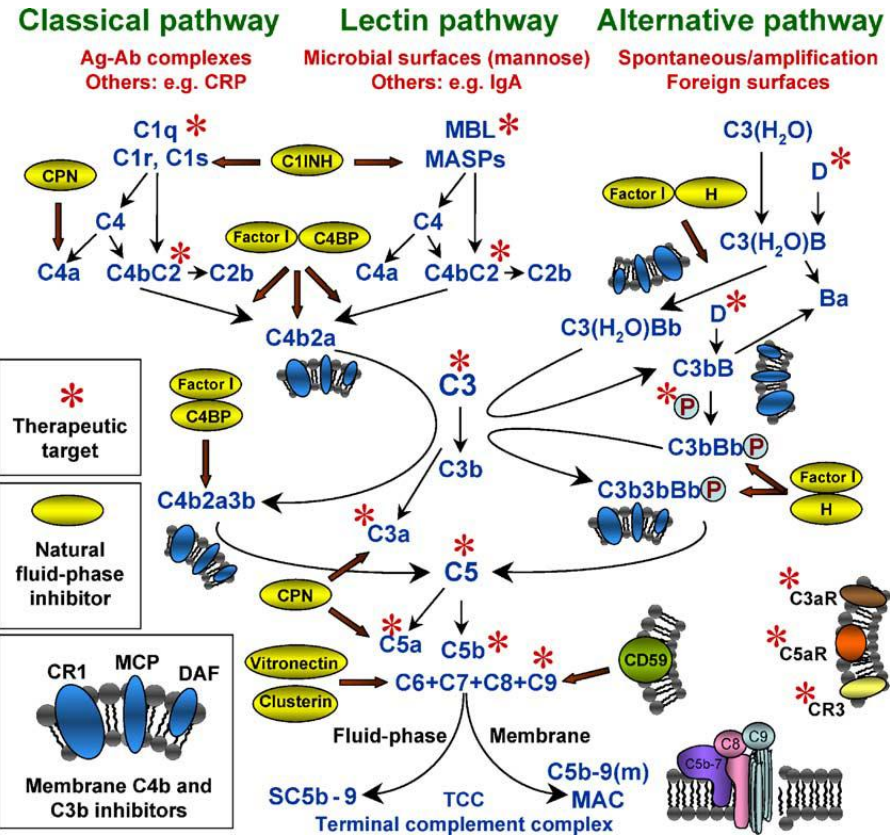
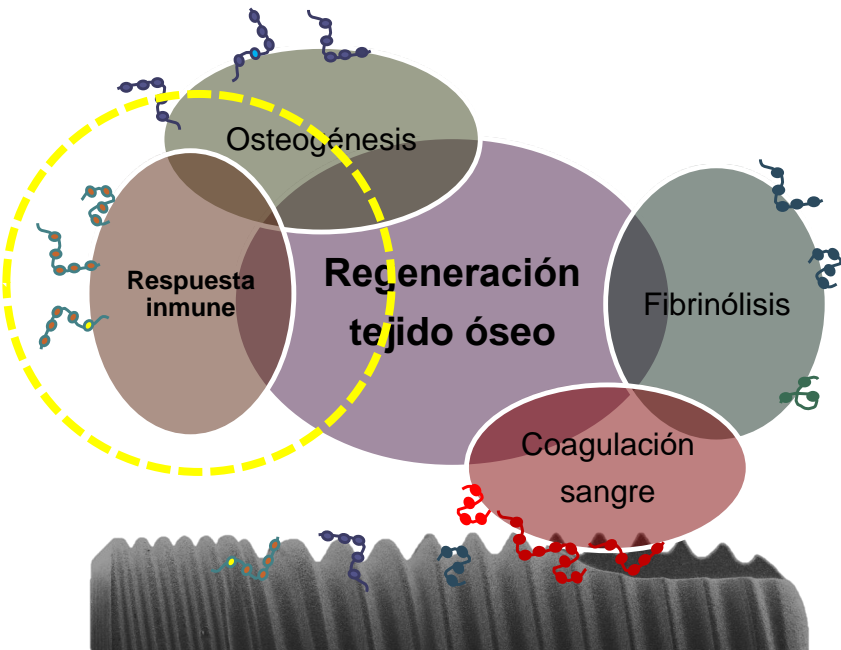
1º: RESPUESTA INMUNE



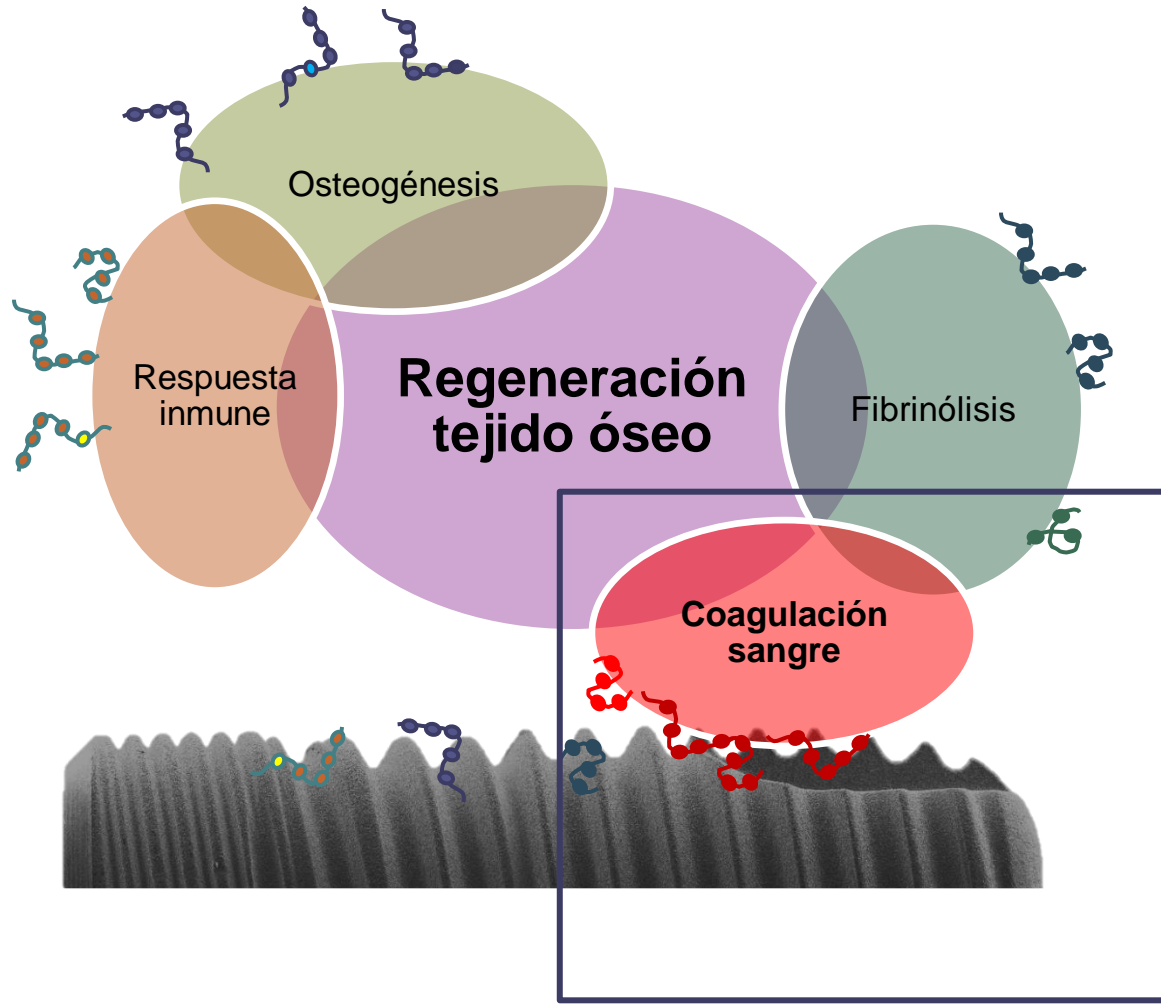
Cascada del sistema del complemento

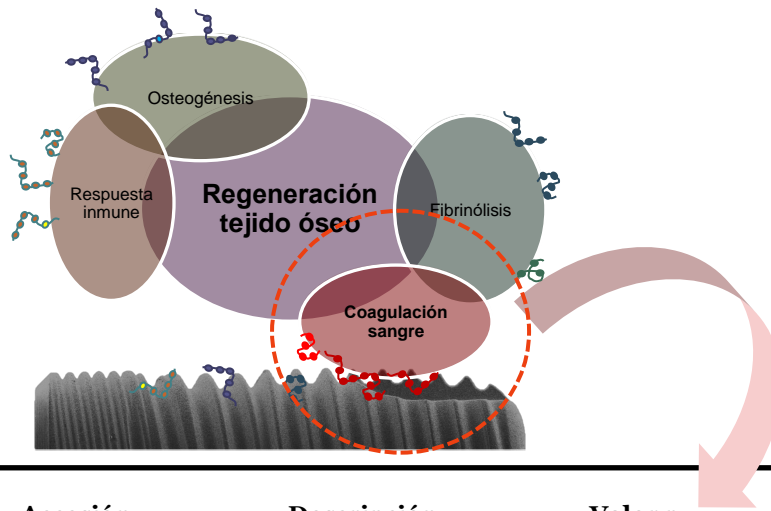
No se detecta una adsorción desproporcionada de proteínas del sistema del complemento, ni en los implantes de GMI, ni en los de Straumann.

No hay proteínas relacionadas con la inflamación crónica (GMI & Straumann)



2º: COAGULACIÓN DE LA SANGRE



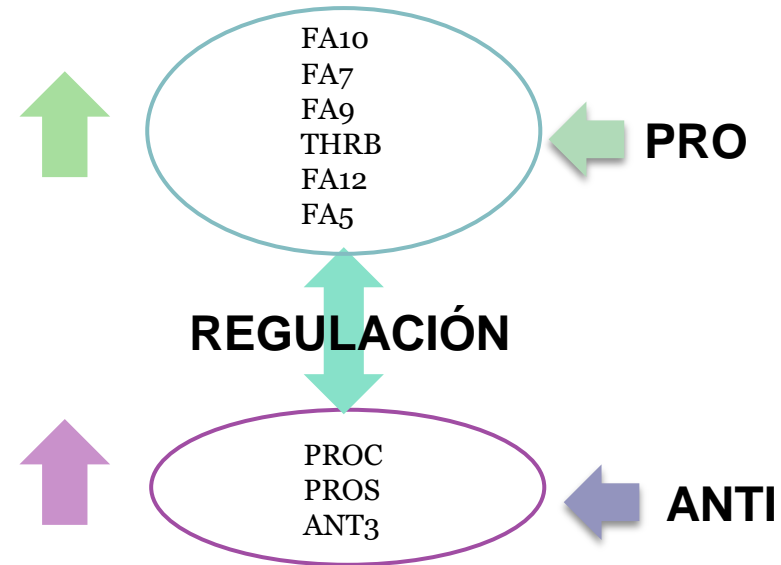


Comparación GMI vs ST

GMI: Alta actividad de coagulación:

Proteínas pro- y anti- coagulantes se adhieren más a la superficie de los implantes de GMI.

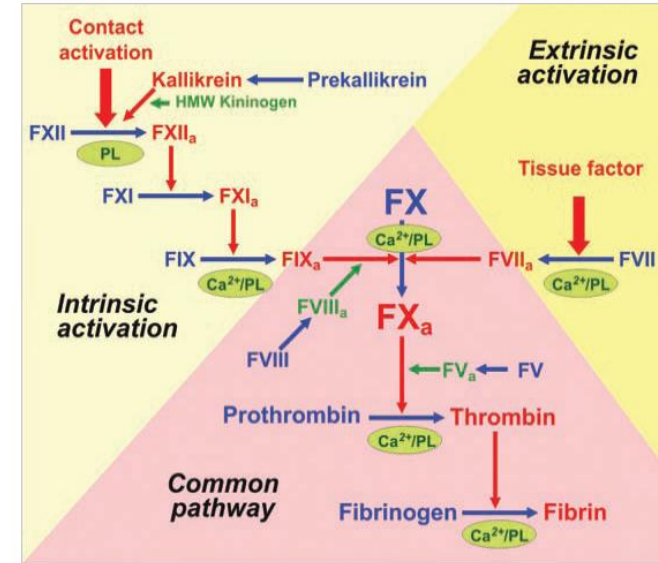
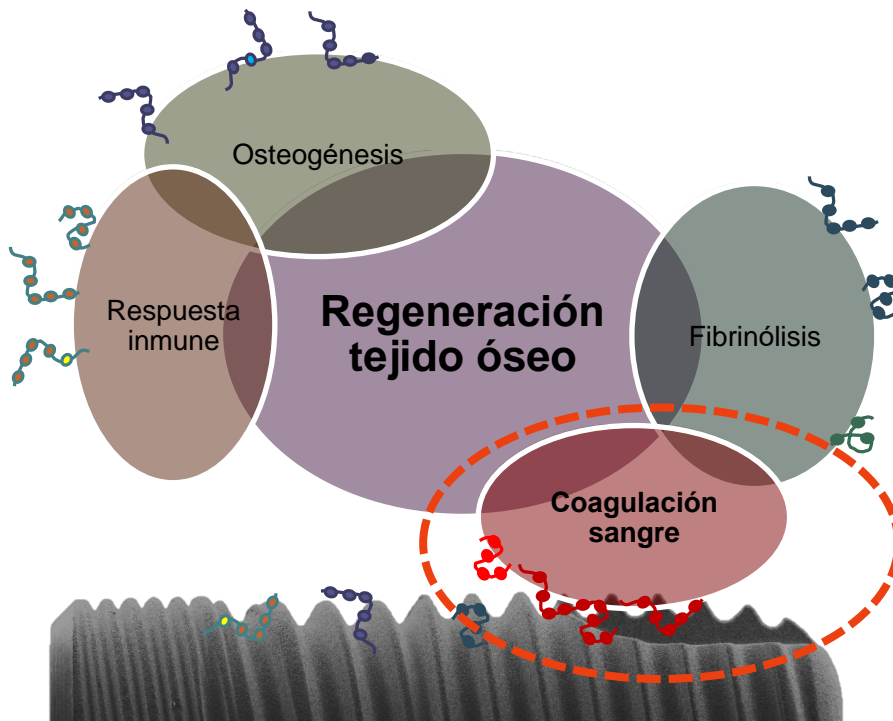
Accesión	Descripción	Valor p	Ratio
PROC_HUMAN	Vitamin K-dependent protein C	1,17E-06	27,21
FA10_HUMAN	Coagulation factor X	1,86E-04	17,21
FA7_HUMAN	Coagulation factor VII	1,04E-04	16,27
FA9_HUMAN	Coagulation factor IX	2,44E-04	5,47
PROS_HUMAN	Vitamin K-dependent protein S	2,07E-03	2,87
ANT3_HUMAN	Antithrombin-III	6,32E-04	2,33
THRB_HUMAN	Prothrombin	9,76E-04	2,31
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FA5_HUMAN	Coagulation factor V	2,80E-02	2,12
KNG1_HUMAN	Kininogen-1	6,98E-05	0,33
FA11_HUMAN	Coagulation factor XI	6,27E-03	0,33
A2MG_HUMAN	Alpha-2-macroglobulin	3,84E-03	0,11
FA11_HUMAN	Coagulation factor XI	6,27E-03	0,33
A2MG_HUMAN	Alpha-2-macroglobulin	3,84E-03	0,11



Los implantes GMI mejoran el proceso de coagulación

Prothrombin

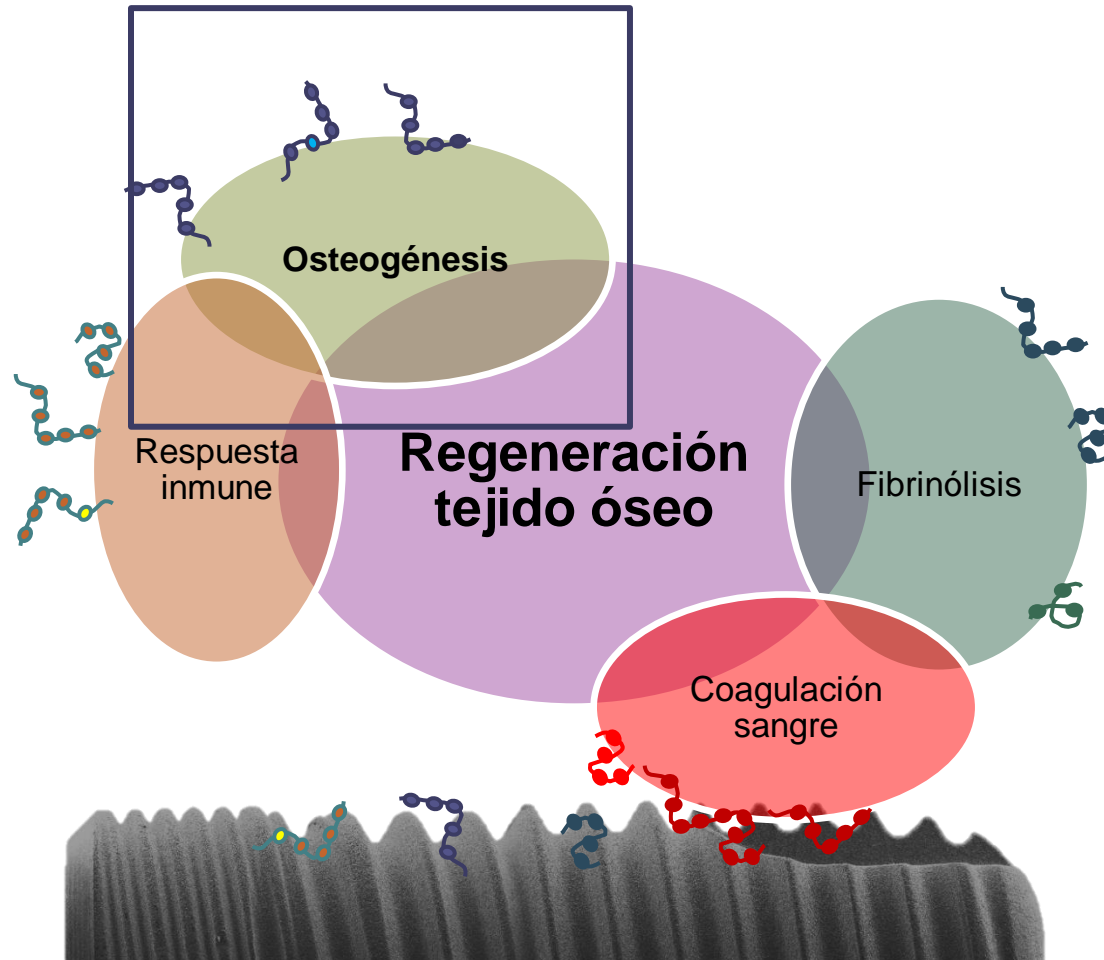
Esta proteína juega un papel clave en la coagulación sanguínea



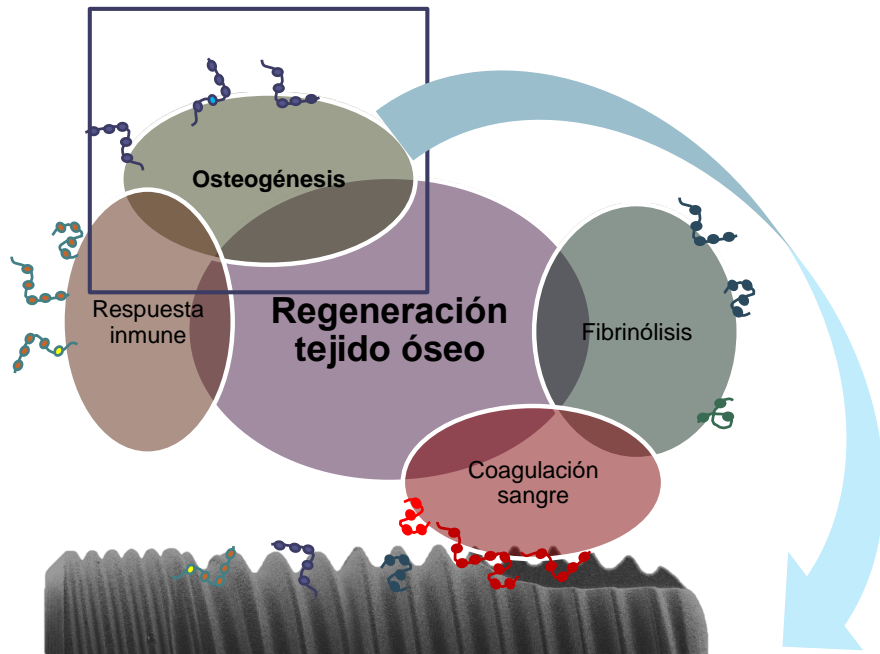
Sin embargo, la coagulación sanguínea es un sistema altamente regulado.

ANT3, PROC, PROS podrían asegurar el desarrollo de un proceso correcto, evitando la inflamación aguda y la trombosis.

3º: OSTEOGÉNESIS



LOS IMPLANTES GMI MEJORAN LA OSTEOGÉNESIS



Mayor afinidad con implantes GMI

GMI



Accesión

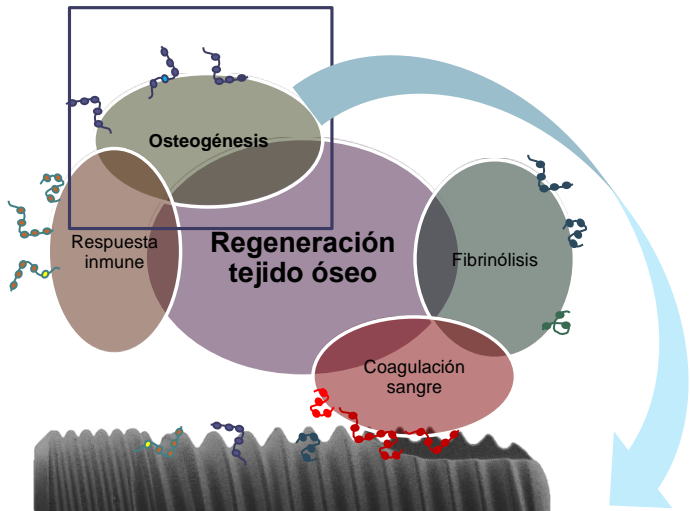
APOE_HUMAN

SEPP1_HUMAN

Description

Apolipoprotein E

Selenoprotein P



Mayor afinidad con implantes GMI

GMI



Accesión	Description
APOE_HUMAN [®]	Apolipoprotein E [®]
SEPP1_HUMAN [®]	Selenoprotein P [®]

Comparación GMI vs ST



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journal homepage: www.elsevier.com/locate/bbagen



Review

Selenoproteins and selenium status in bone physiology and pathology



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ABSTRACT

Background: Emerging evidence supports the view that selenoproteins are essential for maintaining bone health. *Scope of review:* The current state of knowledge concerning selenoproteins and Se status in bone physiology and pathology is summarized.

Major conclusions: Antioxidant selenoproteins including glutathione peroxidase (GPx) and thioredoxin reductase (TrxR), as a whole, play a pivotal role in maintaining bone homeostasis and protecting against bone loss. GPx1, a major antioxidant enzyme in osteoclasts, is up-regulated by estrogen, an endogenous inhibitor of osteoclastogenesis. TrxR1 is an immediate early gene in response to 1 α ,25-dihydroxyvitamin D3, an osteoblastic differentiation agent. The combination of 1 α ,25-dihydroxyvitamin D3 and Se generates a synergistic elevation of TrxR1 activity in Se-deficient osteoblasts. Of particular concern, pleiotropic TrxR1 is implicated in promoting NF κ B activation. Coincidentally, TrxR inhibitors such as curcumin and gold compounds exhibit potent osteoclastogenesis inhibitory activity. Studies in patients with the mutations of selenocysteine insertion sequence-binding protein 2, a key trans-acting factor for the co-translational insertion of selenocysteine into selenoproteins have clearly established a causal link of selenoproteins in bone development. Se transport to bone relies on selenoprotein P. Plasma selenoprotein P concentrations have been found to be positively correlated with bone mineral density in elderly women.

General significance: A full understanding of the role and function of selenoproteins and Se status on bone physiology and pathology may lead to effectively prevent against or modify bone diseases by using Se.



Contents lists available at ScienceDirect

Bone

journal homepage: www.elsevier.com/locate/bone



Original Full Length Article

The role of Apolipoprotein E in bone metabolism

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ABSTRACT

Apolipoprotein E (apoE) is a major structural apolipoprotein of several lipoprotein classes. Over the last 13 years, numerous studies have focused on the question whether human apoE affects bone phenotypes and, more recently, whether apoE regulates bone metabolism in mice. Here, we first provide a brief background introduction into the structure, established physiological and pathophysiological functions of apoE, and will then discuss the new aspects of the emerging role of apoE in bone.

Potencial osteogénico de la proteína ApoE

Aumenta la diferenciación
osteoblástica



Disminuye la diferenciación
osteoclástica



54

BioScience Trends. 2016; 10(1):54-66.

Original Article

DOI: 10.5582/bst.2016.01006

Multifarious effects of 17- β -estradiol on apolipoprotein E receptors gene expression during osteoblast differentiation *in vitro*

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Summary

Apolipoprotein E (ApoE) regulated bone metabolism in mice might mediate uptake of lipid particles into target cells such as osteoblasts *via* receptor-mediated endocytosis by apoE receptors, which includes the low-density lipoprotein receptor (LDLR) family and heparan sulfate proteoglycans (HSPGs). There is no report regarding the expression of ApoE receptors mRNA induced by estrogen during osteoblast differentiation *in vitro*. Primary osteoblasts were collected from the calvaria of newborn mice and were subjected to osteoblast mineralization culture with serial concentrations of 17- β -estradiol (E2) *in vitro*. RNA was isolated at days 0, 5 and 25 of differentiation. Real-time PCR was conducted to analyze apoE receptors mRNA levels. We found that most LDLR family members genes were induced during osteoblast differentiation *in vitro*. The effect of E2 on apoE receptors gene expression during osteoblast differentiation was multifarious. The most noted members of the LDLR family involved in the maintenance of bone metabolism were LRP5, LRP6, LRP4, and Apoer2. LRP6 was up-regulated, while LRP5, LRP4, and Apoer2 were down-regulated by E2. Given that LRP6 is required for early stages of differentiation, we speculate E2 promotes osteoblast differentiation mainly in the early stage.

Keywords: 17- β -estradiol, Apolipoprotein E receptors, Low-density Lipoprotein Receptors Family, Heparan sulfate proteoglycans, Osteoblast differentiation, Reproductive endocrine metabolic network



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journal homepage: www.elsevier.com/locate/yexcr



Research Article

Apolipoprotein E inhibits osteoclast differentiation via regulation of c-Fos, NFATc1 and NF- κ B

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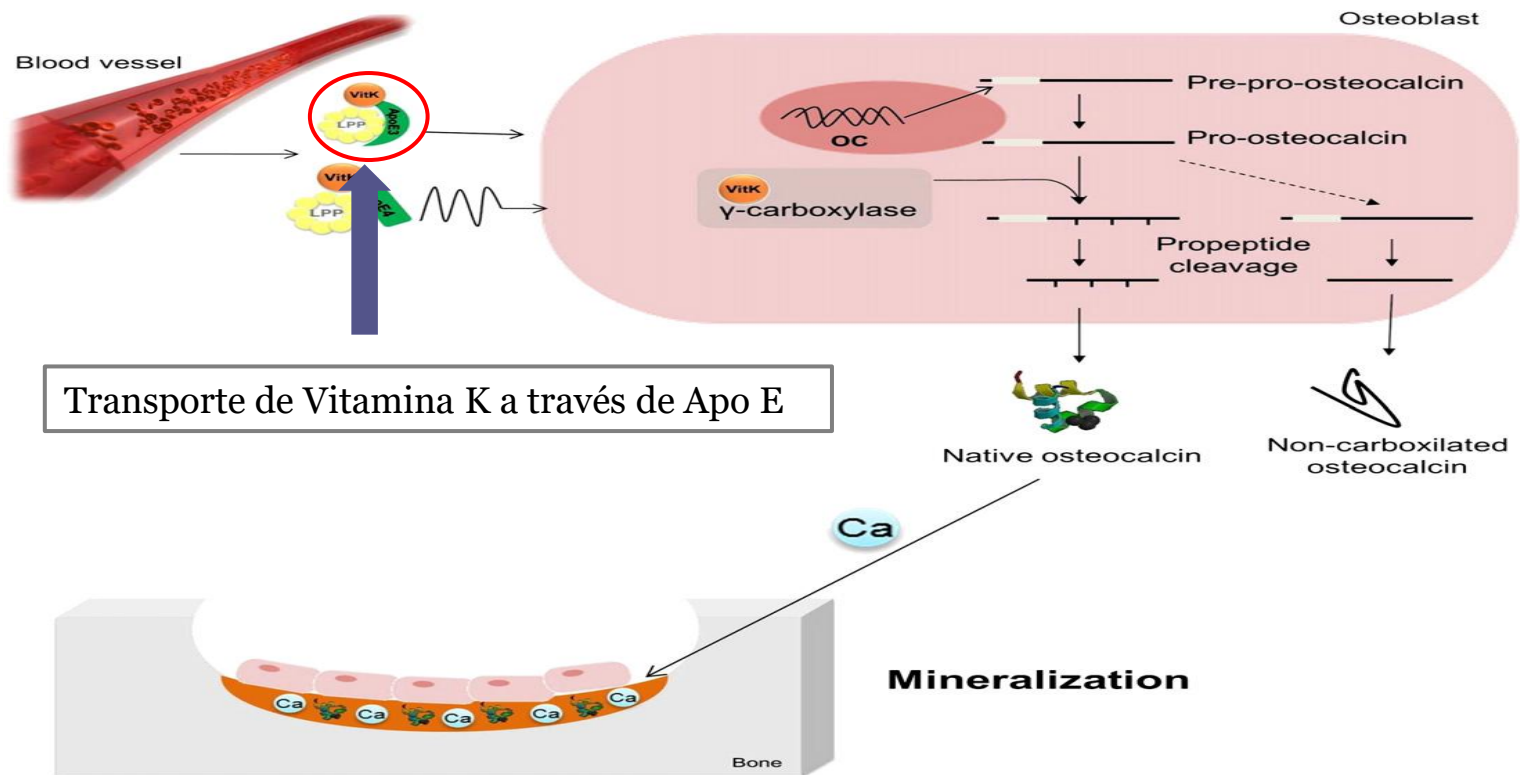
NFATc1

NF- κ B

ABSTRACT

Apolipoprotein E (ApoE) plays a major role in the transport and metabolism of lipid. Other functions of ApoE include modulation of innate and adaptive immune responses. The expression of ApoE in osteoblasts and its relevance with bone formation have also been reported. However, the effect of ApoE on osteoclasts has not yet been examined. Here, we investigated the role of ApoE in osteoclast differentiation using bone marrow-derived macrophages (BMMs) and RAW264.7 cells. We found a down-regulation of ApoE gene expression during osteoclastic differentiation of those cells. Overexpression of ApoE in BMMs and RAW264.7 cells significantly blocked the induction of c-Fos and nuclear factor of activated T cell c1 (NFATc1), transcription factors critical for expression of osteoclast marker genes, by receptor activator of nuclear factor κ B ligand (RANKL), the osteoclast differentiation factor. ApoE inhibited osteoclast differentiation, as measured by decreased number of tartrate-resistant acid phosphatase (TRAP)-positive multinuclear cells (MNCs). In addition, ApoE reduced the expression of dendritic cell-specific transmembrane protein (DC-STAMP) and ATPase, H⁺ transporting, lysosomal 38 kDa, V0 subunit d2 (ATP6v0d2), genes involved in cell-cell fusion during osteoclastogenesis. Knock-down of ApoE using a specific siRNA promoted the RANKL-mediated induction of osteoclast differentiation. While ApoE did not affect the activation of ERK, JNK, and p38 MAPK signaling pathways by RANKL, the phosphorylation of p65 trans-activation domain on serine 536 and transcription activity of NF- κ B were reduced by ApoE overexpression. These findings suggest that ApoE plays an inhibitory role in osteoclast differentiation via the suppression of RANKL-dependent activation of NF- κ B and induction of c-Fos and NFATc1.

Papel de la vitamina K y la ApoE: importancia en el metabolismo óseo



Rodrigues, A. *et al.* Low osteocalcin/collagen type I bone gene expression ratio is associated with hip fragility fractures. *Bone* 51, 981–9 (2012)

CONCLUSIONES

ACTIVIDAD	GMI	STRAUMMAN
RESPUESTA INMUNE	—	—
COAGULACIÓN SANGRE	↑	↓
OSTEOGÉNESIS	↑	↓